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**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

Applicant : John B. Sullivan  
Serial No : 08/405,454  
Conf. No : 6004  
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For : ANTIVENOM COMPOSITION CONTAINING FAB FRAGMENTS  
(As Amended)  
Examiner : Ronald B. Schwadron  
Art Unit : 1644

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**APPEAL BRIEF UNDER 37 C.F.R. § 1.192 (b)(1)**

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### **Real Party in Interest**

The real party in interest in the pending Appeal is the Assignee, Therapeutic Antibodies, Inc., by virtue of an Assignment from Appellants, duly recorded. Therapeutic Antibodies, Inc. has since merged with Proteus International, Inc. to form Protherics PLC.

### **2. Related Appeals and Interferences**

Appellants, the Undersigned, and the Assignee, know of no pending appeals or interferences that will directly affect or be directly affected by or have a bearing on the Board's decision in this Appeal. As Appellants discuss more fully below, this application was subject to a previous appeal (Appeal No. 2001-1255; "the Previous Appeal") involving, *inter alia*, a § 103 rejection of previous versions of claims 40-42. In the Previous Appeal, the Board vacated the § 103 rejection of claims 40-42 but entered a new § 103 rejection of claims 40-42 over the same references.

### **3. Status of Claims**

Claims 40-42, 50, and 54-55 are pending. Claims 54-55 stand withdrawn from consideration by the Examiner.<sup>1</sup> Claims 40-42 and 50 stand rejected by the Examiner under 35 U.S.C. § 103(a). The Appendix contains the pending claims on appeal—claims 40-42 and 50.

### **4. Status of Amendments**

Appellants are concurrently filing an Amendment after Final Office Action requesting the cancellation of claims 51-53 and the amendment of claim 54 to reflect the cancellation of those claims. Because the Amendment After Final Office Action merely cancels claims and removes

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<sup>1</sup> Claims 54-55 are method claims that multiply depend from product claims 40-42 and 50. Appellants would be entitled to present such claims after an indication of allowability of the product claims from which they depend. M.P.E.P. § 821.04 Rejoinder ("Where the application as originally filed discloses the product and the process for making and/or using the product, and only claims directed to the product are presented for examination, when a product claim is found allowable, applicant may present claims directed to the process of making and/or using the patentable product by way of amendment pursuant to 37 C.F.R. § 1.121.") As suggested by the MPEP, however, Appellants have already presented these claims rather than present them after an indication of allowability of the product claims. *See id.* ("In view of the rejoinder procedure, and in order to expedite prosecution, Appellants are encouraged to present such process claims, preferably as dependent claims, in the application at an early stage of prosecution. ")

issues from appeal it will be entered. 37 C.F.R. § 1.116(b); M.P.E.P. § 714.13. The Appendix reflects these amendments, and contains the resulting pending claims on appeal.

## 5. Summary of Invention

The claimed invention relates to Fab fragments that bind specifically to a venom of a snake of the *Crotalus* genus and that are essentially free from contaminating Fc as determined by immunoelectrophoresis, using an anti-Fc antibody. The claimed invention also relates to an antivenom composition comprising these Fab fragments (claim 40). The source of the Fab fragments can be Ig(G)T (claim 41), the Ig(G)T can be covalent (claim 42), and the Fab can be equine (claim 50).

An antivenom is a suspension of venom neutralizing antibodies that are prepared from the serum of animals (typically, horses) that are hyperimmunized against a specific venom or venoms. [Specification at p. 4, lines 19-22.] Typically, animals are repeatedly injected with increasing doses of venom, and the animals' sera are collected and used to obtain antibodies that can neutralize the venom. Antivenoms are typically used to treat human snake bite victims.

[See, *Id.* at p. 23, lines 1-3.]

An antibody molecule is commonly referred to as an immunoglobulin (Ig) and is shaped like a "Y". [*Id.* at p. 23, lines 25-27.] The two tops of this "Y" contain the two antigen-binding sites of the antibody molecule. Exposing an antibody molecule to the enzyme pepsin results in the two upper arms of this "Y" splitting from the stem of the molecule but remaining attached to each other. This results in one  $F(ab)_2$  fragment, the two upper arms—and their two antigen-binding sites—attached to each other, and an Fc fragment, the stem. [*Id.* at lines 27-31.] Exposing an antibody molecule to the enzyme papain results in the two upper arms of this "Y" splitting both from the stem of the molecule and from each other. [*Id.* at lines 41-43.] This results in two separate Fab fragments (the two upper arms)—thus, two separate antigen-binding sites—and an Fc fragment (the stem). [*Id.* at lines 27-31.] The desired fragments can then be purified by using an affinity column that separates the Fab fragments from the Fc fragments. [*Id.* at p. 7, lines 1-14; p. 8, line 37 to p. 9, line 4; Fig. 6; Fig. 8].

## 6. Issues

Whether the Examiner properly made the following rejections:

A. rejection of claims 40-42 and 50 under 35 U.S.C. § 103(a) as being unpatentable over Sullivan *et al.* in view of Coulter *et al.* [Paper No. 48 at p. 4 (item 5)] and

B. rejection of claims 40-42 and 50 under 35 U.S.C. § 103(a) as being unpatentable over Sullivan *et al.* in view of Coulter *et al.* and Smith *et al.* as evidenced by Stedman's Medical Dictionary [Paper No. 48 at p. 5 (item 9)].

## 7. Grouping of Claims

Claims 40-42 and 50 stand or fall together concerning both the rejection under 35 U.S.C. § 103(a) over Sullivan *et al.* in view of Coutler *et al.* and the rejection under 35 U.S.C. § 103 over Sullivan *et al.* in view of Coutler *et al.* and Smith *et al.* as evidenced by Stedman's Medical Dictionary for purposes of this Appeal only.

## 8. Argument

The issues in this appeal are closely related to those from the Previous Appeal. Indeed, they arise directly from the Board's decision in the Previous Appeal. The Board's decision in the Previous Appeal regarding the two § 103 rejections over Sullivan *et al.* in view of Coulter *et al.* turned on whether or not the claims required a pharmacological activity. Thus, the Board affirmed the § 103 rejections of claims 45-47, which were directed to Fab fragments, but the Board vacated the § 103 rejections of claims 40-42, which differed from claims 45-47 only by reciting that the claimed composition is an "antivenom composition."

While the Board vacated the rejection of claims 40-42, it believed "both the examiner and appellants place far too great a weight on the term 'antivenom' in the preamble of [the] claimed composition." [Paper No. 46 at p. 7.] Thus, the Board entered a new ground of rejection for claims 40-42 under 35 U.S.C. § 103 as allegedly being unpatentable over Sullivan *et al.* in view of Coulter *et al.*. [Paper No. 46 at pp. 9-10.] Specifically, the Board asserted that the combination of Sullivan *et al.* in view of Coulter *et al.* taught all the elements of claim 40 except for the pharmaceutically acceptable carrier, which the Board found in Coulter *et al.*'s use of phosphate buffered saline. [Paper No. 45 at pp. 9-10.]

The new ground of rejection was based upon the Board's belief that the mere recitation of an "antivenom composition" in the preamble did not result in the claims requiring a pharmaceutical activity. "There is no requirement in this claim that the Fab fragments exhibit a

pharmaceutical activity.” [Paper No. 45 at p. 9.] Because the Board did not believe claims 40-42 required a pharmaceutical activity, they concluded one of ordinary skill in the art would have combined Sullivan *et al.*’s IgG antivenom teachings with Coulter *et al.*’s teaching that Fab fragments improve the sensitivity of enzyme immunoassays (EIAs) “for use in EIAs to detect said venom.” [Paper No. 46 at p. 9.] In other words, the Board believed that the combination of Sullivan *et al.* and Coulter *et al.* would have suggested using Fab antivenom fragments to detect *Crotalus* venom.

Appellants subsequently amended the preamble of claim 40 to expressly recite that the antivenom composition is an “antivenom pharmaceutical composition for treating a snakebite victim.” Moreover, Appellants also amended the body of claim 40 to recite that the antivenom pharmaceutical composition “neutralizes the lethality of the venom of a snake of the *Crotalus* genus.” These amendments are illustrated below:

40. (Previously amended) An antivenom pharmaceutical composition for treating a snakebite victim, comprising Fab fragments which bind specifically to a venom of a snake of the *Crotalus* genus and which are essentially free from contaminating Fc as determined by immunoelectrophoresis using anti-Fc antibodies, and a pharmaceutically acceptable carrier, wherein said antivenom pharmaceutical composition neutralizes the lethality of the venom of a snake of the *Crotalus* genus.

Thus, there can be no doubt that this claim (as well as the remaining claims, which are similarly amended), now contains a “requirement . . . that the Fab fragments exhibit a pharmaceutical activity.” [Paper No. 45 at p. 9.] The very basis of the Board’s new ground of rejection in the Previous Appeal has therefore been removed.

Despite these amendments to claim 40 to specifically require that the antivenom pharmaceutical composition exhibits a pharmaceutical activity (neutralizes the lethality of the venom of a snake of the *Crotalus* genus), the Examiner maintained the rejection. In doing so, the Examiner merely asserted that the amended preamble was no more limiting than the old preamble. [Paper No. 48 at p. 4.] The Examiner did not even address the clause reciting that the antivenom pharmaceutical composition “neutralizes the lethality of the venom of a snake of the *Crotalus* genus.” Indeed, even in the second rejection under appeal, where the Examiner cited further secondary references, the Examiner made no new arguments. Rather, the Examiner merely referred to the Examiner’s Answer in the Previous Appeal. [Paper No. 48 at p. 6.]

Appellants emphasize that this is not a case of merely stating a new use for an obvious composition, which the Board in the Previous Appeal reminded the examiner and Appellants does not render composition claims patentable. [Paper No. 46 at pp. 7, 9 (citing *In re Zierden*, 162 USPQ 102, 104 (CCPA 1969) and *In re Pearson*, 181 USPQ 641, 644 (CCPA 1974).] Rather, it is a case of the claims reciting terms that “define . . . some characteristic not found in the old composition.” *Pearson*, 181 USPQ at 644. Such terms can “be used to distinguish a new from an old composition.” [*Id.*]

As Appellants discuss in detail below, amended claim 40, as well as the remaining product claims, now recite such terms that define characteristics that patentably distinguish the claimed antivenom pharmaceutical composition from the antivenom composition of *Sullivan et al.* combined with the EIA detection reagent of *Coulter et al.* There was no suggestion in the prior art that Fab fragments could be used to create an antivenom pharmaceutical composition for treating a snakebite victim that would neutralize the lethality of the venom of a snake of the *Crotalus* genus. This lethality neutralization is not just an intended use of the claimed product. It is a required characteristic of the claimed product that distinguishes it from the prior art, which believed that Fab antivenom compositions would not only fail to neutralize the lethality of the venom of a snake of the *Crotalus* genus, but might actually enhance the lethality. Accordingly, both rejections of claims 40-42 and 50 under 35 U.S.C. § 103(a) over *Sullivan et al.* in view of *Coutler et al.* as primary references should be reversed

#### **1. Before Appellants' Invention, Antivenoms Comprising Fab Fragments Were Expected to Be Ineffective in Neutralizing the Lethality of the Venom of a Snake of the *Crotalus* Genus**

Appellants have submitted numerous references and the Declarations of Dr. Damon Smith, Dr. John B. Sullivan, and Findley E. Russell, M.D., Ph.D.<sup>2</sup>, which prove that, at the time of Appellants' invention, the claimed invention would not have been obvious because one of ordinary skill in the art would not have had a reasonable expectation of success. Indeed, Dr. Sullivan "and others questioned whether anti-venom F(ab)'s would be effective [antivenoms]" [Sullivan Decl. at ¶9], and Dr. Sullivan and others actually believed that Fab fragments would

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<sup>2</sup> Dr. John B. Sullivan, and Findley E. Russell, M.D., Ph.D are the named inventors, and they are the authors of the Sullivan et al. reference.

not only fail to neutralize *crotalus* venom, they might actually "increase toxicity of the venom."  
[Sullivan Decl. at ¶ 13; emphasis in original.]

The only commercially available antivenom at the time of Appellants' invention for North American snakes of the *Crotalus* genus was Antivenin [Crotalidae] Polyvalent (equine origin) ("ACP"), which first became available in 1947. [First Russell Decl. at ¶ 20; Smith Decl. at ¶ 7.] This antivenom suffered the serious problem suffered by other antivenoms of often causing serum sickness, an allergic reaction to the antivenom that is sometimes as dangerous as the venom. [First Russell Decl. at ¶ 20; specification at p. 4, lines 35-40.] Over 75% of envenomation patients who receive ACP suffer from serum sickness. [First Russell Decl. at ¶ 20.] This danger can be so great that physicians may not administer this antivenom for some cases of envenomation, and ACP can only be obtained in a kit that also contains test serum for attempting to detect serum sickness. *[Id.]*

Because of the serious problem of serum sickness, extensive research had been performed on developing better antivenoms. [First Russell Decl. at ¶ 24.] It was generally believed that "given possession of the antibody active site, the smaller the antibody molecule, the better. [Specification at p. 3, lines 1-2.] Thus, much of this research focused on immunoglobulin fragments, which may not provoke an immune reaction. [First Russell Decl. at ¶ 24.] In the late 1960's, researchers began experimenting with antivenoms comprising  $F(ab)_2$  fragments, and such antivenoms first became commercially available in 1969. *[Id.;* Smith Decl. at ¶ 17.] Although the smaller size of the  $F(ab)_2$  fragments results in less serum sickness, such antivenoms appeared less effective than antivenoms comprising whole immunoglobulin. [First Russell Decl. at ¶ 25.] Consequently, Crotalidae antivenoms comprising  $F(ab)_2$  fragments were not produced in the United States. [First Russell Decl. at ¶ 24].

Although serum sickness had long been recognized as a major problem with antivenoms, and although smaller antibody fragments had long been known to be less immunogenic, no researcher developed antivenoms comprising the smaller Fab fragments prior to Appellants' invention. *[Id. at ¶ 25; Sullivan Decl. at ¶ 15.]* Indeed, there had been no significant improvements in commercial antivenoms since 1969, when an  $F(ab)_2$  antivenom was commercially sold. [Smith Decl. at ¶ 7.] Development of antivenoms comprising antibody fragments halted at the larger  $F(ab)_2$  fragments because the larger  $F(ab)_2$  fragments appeared to

some of ordinary skill in the art to be less effective than whole antibody. [First Russell Decl. at ¶ 25.]

Not only did the F(ab)<sub>2</sub> fragments, which are larger than Fab fragments, appear to be less effective than whole antibody molecules, but those of ordinary skill in the art expected Fab fragments to be even less effective than the disappointing F(ab)<sub>2</sub> fragments for several reasons. [First Russell Decl. at ¶ 26; Sullivan Decl. at ¶ 5; Smith Decl. at ¶ 9.] First, Fab fragments cannot sterically hinder the binding of a venom protein to its tissue target as well as F(ab)<sub>2</sub> fragments because Fab fragments have only one active site. [First Russell Decl. at ¶ 29; Sullivan Decl. at ¶ 8.] The two binding sites on F(ab)<sub>2</sub> fragments allow them to bind to repeating antigenic determinants on a venom antigen, and this repetitive determinant binding sterically hinders the venom antigen from binding to its active site.

Second, since F(ab)<sub>2</sub> fragments contain two antigen binding sites, each individual F(ab)<sub>2</sub> fragment can bind two antigens. [Steward Sell, *Basic Immunology: Immune Mechanisms in Health and Disease*, at p. 89, Fig. 6-3 (1987).] As more F(ab)<sub>2</sub> fragments cross-link more antigens, they form larger complexes which, eventually, become large enough that they precipitate from solution. *Id.* In contrast, since Fab fragments have only one antigen binding site, they cannot form cross-linked complexes and precipitate the antigens. [Smith Decl. at ¶ 9.]

Third, those of ordinary skill in the art expected that Fab fragments would not be effective because they would be cleared before the venom. Many venom toxins are large, hydrophobic molecules, and they are usually injected deep into subcutaneous tissues. [First Russell Decl. at ¶ 30; Smith Decl. at ¶ 8.] These individual toxins are released slowly from the injection site, resulting in the "venom depot effect" whereby the venom toxins continue to be released into the circulatory system long after the initial bite. [First Russell Decl. at ¶ 30; Smith Decl. at ¶ 8.] Venom protein continues to be released from the injection site for weeks [Sullivan Decl. at ¶ 5(a)], and has been detected in a patient 46 days after envenomation. [Owenby *et al.*, *Southern Medical Journal* (1990).]

Fab fragments have a molecular weight of around 45-55 Kd. [First Russell Decl. at ¶ 31.] This relatively small size allows the renal system to remove Fab fragments, resulting in a half-life of about 17 hours. [*Id.*] Indeed, the renal system completely eliminates Fab fragments in only 24-26 hours. [*Id.*]

F(ab)<sub>2</sub> fragments, in contrast, are about twice as large as Fab fragments-too large for the renal system to remove them. [Id. at ¶ 32.] Thus, they have a much longer half-life than Fab fragments, approximately 50 hours versus approximately 17 hours. [Id.] Given the renal system's rapid removal of Fab fragments, especially compared to F(ab)<sub>2</sub> fragments, and the venom depot effect, those of ordinary skill in the art expected that there would be no remaining Fab fragments to neutralize later-released venom toxins. [Id. at ¶ 32; Smith Decl. at ¶ 8.]

For all these reasons, antivenoms comprising Fab fragments were expected to be ineffective in neutralizing the lethality of the venom of a snake of the *Crotalus* genus. Thus, despite the long-known, serious deficiencies of the only commercially available antivenom, no researchers developed antivenoms comprising the smaller Fab fragments prior to Appellants' invention.

**2. Not Only Were Antivenoms Comprising Fab Fragments Expected to Be Ineffective in Neutralizing the Lethality of the Venom of a Snake of the *Crotalus* Genus, they Were Actually Expected to Be Harmful**

Not only did those of ordinary in the skill in the art believe that Fab fragments would be ineffective in neutralizing the lethality of the venom of a snake of the *Crotalus* genus, they actually expected that such an antivenom could increase the lethality of the snake venom by redistributing and concentrating its toxins. [Russell Decl. at ¶ 33; Sullivan Decl. at ¶ 13.] The binding of Fab fragments and venom toxins is a dynamic process, having an equilibrium where individual venom toxins are constantly bound and released. [First Russell Decl. at ¶ 34.] The renal system's rapid removal of Fab fragments, however, continually decreases the number of Fab fragments remaining to bind the venom toxins. [Smith Decl. at ¶ 8.] Those of ordinary skill in the art were concerned that Fab fragments would bind venom toxins that were released into the circulatory system and then release the venom toxins at another site, perhaps concentrating the venom toxins in areas of high blood flow like the kidneys, heart, nervous system, and lungs. [First Russell Decl. at ¶ 36; Sullivan Decl. at ¶ 5(b).] As Dr. Sullivan stated,

I and others maintained and discussed our concerns that Fab[fragments] would redistribute toxic venom proteins throughout the body, thus producing venom pathology at tissue sites and organ systems not typically seen in patients treated with [whole antibodies] or F(ab)<sub>2</sub>.

[Sullivan Decl. at ¶ 17.] While the toxins might have caused swelling and local necrosis at the site of envenomation, the predicted redistribution and concentration of venom toxins might result in "coagulopathy, direct cardiotoxicity, liver and kidney damage, potential central nervous system, and peripheral nervous system damage." [Id.] Thus, what had been a systemic toxicity with venom toxins being released slowly into the circulation could become a localized toxicity with venom toxins being concentrated in the kidneys, heart, nervous system, and lungs by this "taxi" effect. [First Russell Decl. at ¶ 36; Sullivan Decl. at ¶ 5(a).]

This taxi effect was predicted, and it was a reason why those of ordinary skill in the art did not progress beyond the known  $F(ab)_2$  fragments to the smaller Fab fragments. [Sullivan Decl. at ¶ 7.] According to Dr. Sullivan, the use of Fab fragments to treat envenomation would have been "medically unsound and contraindicated." [Id. at ¶ 13.] The belief of those of ordinary skill in the art that Fab fragments would actually increase the lethality of snake venom by concentrating high molecular weight snake toxins in areas of high blood flow was not a merely theoretical concern, as Faulstich *et al.* later demonstrated.

Faulstich *et al.* (Strongly Enhanced Toxicity of the Mushroom Toxin  $\alpha$ -Amanitin by an Amatoxin-Specific Fab or Monoclonal Antibody. 26 *Toxicon* 491 (1988) (copy attached as Exhibit 7 to first Russell Declaration)] conducted a series of studies attempting to treat  $\alpha$ -amatoxin poisoning with Fab fragments. Alpha-amatoxin is a high molecular weight toxin that is similar to some snake venom toxins. [First Russell Decl. at ¶ 37.] As a high molecular weight toxin,  $\alpha$ -amatoxin cannot be cleared by the renal system. [Id.] Rather, like many snake toxins, it is cleared by the liver. [Id.] Since  $\alpha$ -amatoxin is concentrated in the liver after oral ingestion, it is primarily toxic to liver cells. [Id.]

Faulstich *et al.* discovered that the Fab fragments did not decrease the toxicity of  $\alpha$ -amatoxin in mice, but rather increased the toxicity of  $\alpha$ -amatoxin by a factor of 50. [Faulstich *et al.* at p. 497.] Furthermore, the Fab fragments resulted in  $\alpha$ -amatoxin being specifically toxic to kidney cells rather than liver cells. [Id.] This is exactly what one of ordinary skill in the art would have predicted. [First Russell Decl. at ¶ 38.] The Fab fragments bound the high molecular weight  $\alpha$ -amatoxin, and then unbound it in their state of equilibrium at sites of high blood flow. [Id.] This unbinding at sites of high blood flow, especially the kidneys, resulted in the  $\alpha$ -amatoxin being concentrated in these tissues and killing them. [Id.] Thus, Fab fragments

greatly increased the toxicity of this high molecular weight toxin by concentrating it in areas of high blood flow.

Faulstich *et al.*'s results with Fab fragment directed to a high molecular weight toxin stood in contrast to Balthazar *et al.*'s results with Fab fragments to the low molecular weight toxin digoxin. [Balthazar *et al.* (1994) Utilization of Antidrug Antibody Fragments for the Optimization of Intraperitoneal Drug Therapy: Studies Using Digoxin as a Model Drug. *J. Pharm. Exp. Ther.* 268, 734 (attached at Exhibit 8 to the First Russell Declaration).] Digoxin is unlike most Crotalidae venom toxins; it is a very small molecule; small enough that the renal system can clear the Fab-digoxin complex. [First Russell Decl. at ¶ 39; Smith Decl. at ¶¶ 8, 10.] Since the renal system can filter the Fab-digoxin complex, the Fab fragments did not redistribute and concentrate digoxin, as one of ordinary skill in the art would have predicted. [First Russell Decl. at ¶ 39.] Accordingly, Balthazar *et al.* found that Fab fragments effectively treated digoxin toxicity, just as Smith *et al.*, upon which the Examiner relies, found.

However, Balthazar *et al.* recognized the potential problems of Fab therapy for large toxins, like  $\alpha$ -amatoxin and some Crotalidae venom toxins:

First, the alteration of drug distribution which accompanies antibody drug complexation may result in a potentiation of drug toxicities or the development of new drug toxicities in certain cases . . . The risk of redistributing systemic toxicity, rather than minimizing systemic toxicity, should be appreciated as a potential outcome of the proposed approach.

[Balthazar *et al.* at p. 738, cols. 1-2; emphasis added.]

Accordingly, those of ordinary skill in the art were concerned that treatment with an antivenom comprising Fab fragments would actually be harmful for the treatment of high molecular weight venom toxins because the Fab fragments would redistribute high molecular weight toxins to areas of high blood flow, creating new toxicities. Faulstich *et al.* confirmed this concern with a toxin that is of a similar molecular weight as many snake venom toxins. [First Russell Decl. at ¶ 41.] Balthazar *et al.* reinforced this concern by showing that this effect did not occur with a low molecular weight toxin that the renal system could clear as part of an Fab-toxin complex. [First Russell Decl. at ¶ 42.] Indeed, despite the effectiveness of their treatment, Balthazar *et al.* specifically discussed their concern that Fab fragments might alter drug toxicities or redistribute systemic toxicities.

3. **One of Ordinary Skill in the Art Would Not Have Extrapolated Coulter et al.'s *in vitro* Results with the Single Venom Toxin Textilotoxin to the *in vivo* Neutralization of the Lethality of a Whole Venom**

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Against this evidence that those of ordinary skill in the art would not have expected an antivenom pharmaceutical composition comprising Fab fragments to actually neutralize the lethality of the venom of a snake of the *Crotalus* genus, the Examiner has relied upon Coulter *et al.* for teaching the benefits of Fab fragments as antitoxins in general. One of ordinary skill in the art would not have been motivated with a reasonable expectation of success to make the claimed invention for the further reason that they would not have extrapolated Coulter *et al.*'s *in vitro* results with a single venom toxin to the *in vivo* neutralization of the lethality of a whole venom.

The same *in vivo* mechanisms that led those of ordinary skill in the art to expect that the claimed invention might actually increase the lethality of the venom of a snake of the *Crotalus* genus show that the Examiner's reliance upon the Coulter *et al.* reference for teaching that Fab fragments have a higher sensitivity than whole antibody *in vitro* tests is misplaced. Coulter *et al.* did not treat envenomation with their Fab fragments. Rather, Coulter *et al.* first mixed textilotoxin with their Fab fragments *in vitro*. [Coulter *et al.* at p. 201, 3rd full paragraph.] Coulter *et al.* then injected the already bound Fab-textilotoxin complex intravenously. This treatment with Fab fragments resulted in neutralization that was essentially equivalent to the treatment with the IgG fragments, just as one of ordinary skill in the art would have expected. [First Russell Decl. at ¶ 48.] Since the Fab-textilotoxin mixture was first mixed *in vitro* and then injected intravenously, the Fab did not have the opportunity to redistribute and concentrate the textilotoxin in high blood flow parts. [*Id.*] Accordingly, the Coulter *et al.* reference would not have provided a reasonable expectation of success for an antivenom comprising Fab fragments to any venom toxins, despite the Examiner's assertion to the contrary. [*Id.*]

Once again, this was not a merely theoretical concern, as Sorkine *et al.* later demonstrated. Sorkine *et al.* conducted a similar experiment in 1983 by mixing Fab fragments with a snake venom before injecting the mixture into a mouse, and they obtained similar results. [Sorkine *et al.* (1995) Comparison of F(ab')<sub>2</sub> and Fab Efficiency on Plasma Extravasation Induced *Viper aspis* Venom. *Toxicon* 33, 257 (attached as Exhibit 11 to the First Russell

Declaration).] This treatment resulted in a considerable reduction in capillary permeability. However, the Fab fragments were much less effective when they were administered *in vivo* separately from the venom. As Sorkine *et al.* state "these data showed firstly that the *in vitro* neutralization of the venom by immunoglobulin fragments does not reflect their *in vivo* efficiency." [Sorkine *et al.* at 257.] Thus, the Sorkine *et al.* reference shows that one would not have expected Coulter *et al.*'s *in vitro* neutralization results to predict the effectiveness of antivenoms comprising Fab fragments *in vivo*. [First Russell Decl. at ¶ 150.]

Moreover, Coulter's results Coulter *et al.* used textilotoxin, one of several toxins in the venom of the Australian brown snake (*Pseudonaja textilis*). [Coulter *et al.* at p. 199, last sentence; First Russell Decl. at ¶ 46.] The pending claims recite a snake of the genus *Crotalus*, a genus of the family Crotalidae. As can be seen from its name, the snake Coulter *et al.* used is not a member of the genus *Crotalus*, nor is it even of the same family as the *Crotalus* genus. Rather, it is a member of the genus *Pseudonaja*. [Coulter *et al.* at 199.] Indeed, Coulter *et al.*'s snake is an elapid [Russell (1996) *Toxic Effects of Animal Toxins*. In *Casarett and Doull's Toxicology: The Basic Science of Poisons*, (5th Ed.) at p. 802 (attached as Exhibit 10 to the First Russell Declaration)], and the elapids are of the family Elapidae, not Crotalidae. [Snake Venom Poisoning at p. 5.]

Textilotoxin is simply a single toxin from the venom of a snake of the *Pseudonaja* genus. Although venoms can be simple substances, as in some marine animals, in snakes they are often very complicated mixtures of many individual toxins. [First Russell Decl. at. ¶ 15, ¶ 47; Smith Decl. at ¶ 6.] In some venoms of *Crotalus* snakes, there may be 100 different protein fractions. [First Russell Decl. at ¶ 15.] Due to their complexity, the full composition of snake venoms is unknown. [Id.] Not only is the composition of snake venoms complicated and their exact composition unknown, but the pharmacological effects of some constituent toxins are unknown. [Id. at ¶ 16.]

Due to the unknown composition of snake venoms and the unknown effect of even the identified toxins in snake venoms, basic toxicology texts caution against extrapolating results from individual venom toxins (like Coulter *et al.*'s) to whole venoms (like the claims recite). [*Toxic Effects of Animal Toxins* at p. 802; *Snake Venom Poisoning* at p. 168.] Accordingly, the Examiner is incorrect in attempting to extrapolate Coulter *et al.*'s results with Fab fragments to a single snake venom toxin to the results that would have been expected with Fab fragments to an

entire snake venom comprising many unknown toxins of unknown effect. As Dr. Russell stated, "one would not have expected Coulter *et al.*'s results with Fab to a single **toxin** to predict similar results with Fab to a Crotalidae snake **venom**, including a *Crotalus* snake **venom**." [First Russell Decl. at ¶ 47; emphasis in original.] Since Coulter *et al.* used Fab fragments to a toxin from the venom of a snake of a different genus than the claims recite, and since one of ordinary skill in the art would not have expected results with Fab fragments to a single venom toxin to predict what would occur with an antivenom comprising Fab fragments to an entire *Crotalus* venom, any rejection relying upon the Coulter *et al.* reference must fail.

"Hindsight is not a justifiable basis on which to find" that an invention was obvious. *Amgen v. Chugai Pharm. Co. Ltd.*, 18 U.S.P.Q.2d 1016, 1023 (Fed. Cir. 1991). Rather, the prior art, not Appellants' disclosure, must have provided a reasonable expectation of success. [*Id.*] at 1022; *In re Vaeck*, 20 U.S.P.Q.2d 1438, 1442 (Fed. Cir. 1991). Reliance upon Coulter *et al.*'s *in vitro* results with a single venom toxin of the *Pseudonaja* genus, however, depends upon a hindsight-aided application of the teachings from Appellants' disclosure, not the prior art—one that basic toxicology test caution against making. In contrast to this hindsight, the evidence Appellants have submitted shows that this rejection fails for a lack of the required expectation of success.

In sum, prior to Appellants' invention, those of ordinary skill in the art did not have a reasonable expectation of success that an antivenom pharmaceutical composition comprising Fab fragments to *Crotalus* venom would be effective at neutralizing the lethality of the venom of snake of the *Crotalus* genus. Obviousness requires that "[b]oth the suggestion and the reasonable expectation of success must be founded in the prior art, not [Appellants'] disclosure," *Vaeck*, 20 U.S.P.Q.2d at 1442, and Appellants have shown that that is not the case here. Despite the long-known problems with the commercially available venom for *Crotalus* envenomation since 1947, and the well-known fact that smaller immunoglobulin fragments are less immunogenic, those of ordinary skill in the art had not progressed beyond antivenoms comprising the disappointing F(ab)<sub>2</sub> fragments to the smaller Fab fragments because they expected Fab fragments to be not just ineffective, but actually more harmful to the patient than no treatment at all.

For all these reasons, any combination of Sullivan *et al.* and Coulter *et al.*, including the addition of Smith *et al.* and evidence from Stedman's Medical Dictionary, would not have taught one of ordinary skill in the art to prepare an antivenom pharmaceutical composition for treating a

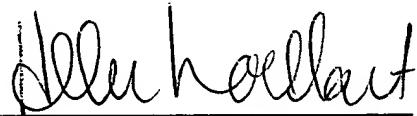
snakebite victim, comprising Fab fragments, wherein the antivenom pharmaceutical composition neutralizes the lethality of the venom of a snake of the *Crotalus* genus.

9. **Conclusion**

In view of the foregoing Appellants respectfully request reversal of the two rejections of claims 40-42 and 50 under 35 U.S.C. § 103 and allowance of the pending claims.

If a further extension of time under 37 C.F.R. § 1.136 is required to obtain entry of this Appeal Brief, such an extension is hereby respectfully requested. If there are any fees due under 37 C.F.R. §§ 1.16 or 1.17 that are not enclosed herewith, including any fees required for an extension of time under 37 C.F.R. § 1.136, please charge such fees to our Deposit Account No. 23/2825.

Respectfully submitted,

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## APPENDIX

40. An antivenom pharmaceutical composition for treating a snakebite victim, comprising Fab fragments which bind specifically to a venom of a snake of the *Crotalus* genus and which are essentially free from contaminating Fc as determined by immunoelectrophoresis using anti-Fc antibodies, and a pharmaceutically acceptable carrier, wherein said antivenom pharmaceutical composition neutralizes the lethality of the venom of a snake of the *Crotalus* genus.

41. The antivenom pharmaceutical composition of claim 40, wherein an antibody source for said Fab fragments is IgG(T).

42. The antivenom pharmaceutical composition of claim 40, wherein an antibody source for said Fab fragments is polyvalent IgG(T).

50. The antivenom pharmaceutical composition of claim 40, wherein the Fab fragments are equine.